Instructions for UseLife Science Kits & Assays





Order No.:

31-DP-1000010 10 reactions 31-DP-1000050 50 reactions 31-DP-1000250 250 reactions



IVD

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1 Introduction

1.1 Intended use

The deltaPREP Blood DNA Mini Kit (MDX) has been designed as a very efficient tool for fast isolation of genomic DNA from whole blood samples for subsequent in vitro diagnostic purposes. The kit can be used for isolation of genomic DNA from fresh or frozen blood; stabilized with EDTA or citrate, from common blood collection systems. Sample volumes from 200 μ l up to 400 μ l can be processed. If smaller volumes of blood are used, apply sterile PBS up to 200 μ l final sample volume.

The extraction procedure is based on a new patented chemistry and combines lysis of blood sample with subsequent binding of nucleic acids onto the surface of a Spin Filter membrane. After several washing steps the nucleic acids are eluted from the membrane by using Elution Buffer. Extraction chemistry and extraction protocol are optimized to get maximum of yield.

The deltaPREP Blood DNA Mini Kit (MDX) is not for use with cell-free body fluids such as cerebrospinal fluid, serum, plasma or urine, tissue or stool samples. The kit performance has not been evaluated with buffy coat, cultured or isolated cells, swabs, dried blood spots and viral DNA. The kit is also not specified for the isolation and purification of fungal, bacterial or parasite nucleic acids.

The kit is intended for use by professional users. The kit has been designed to be used for a wide range of different downstream applications, like amplification reactions and further analytical procedures. Diagnostic results generated using the extraction procedure in conjunction with diagnostic tests should be interpreted with regard to other clinical or laboratory results. To reduce irregularities in diagnostic results, internal controls for downstream applications should be used.

The deltaPREP Blood DNA Mini Kit (MDX) does not provide a diagnostic result. It is the sole responsibility of the user to use and validate the kit in conjunction with a downstream in vitro diagnostic assay.



CONSULT INSTRUCTION FOR USE

This package insert must be read carefully prior to use. Package insert instructions must be followed accordingly. Reliability of results cannot be guaranteed if there are any deviations from the instructions in this package insert.

1.2 Notes on the use of this manual

For easy reference and orientation, the manual uses the following warning and information symbols as well as the shown methodology:

Symbol	Information
REF	REF Catalogue number.
\sum_{N}	Content Contains sufficient reagents for <n> reactions.</n>
15℃ Å 30°C	Storage conditions Store at room temperature or shown conditions respectively.
[]i	Consult instructions for use This information must be observed to avoid improper use of the kit and the kit components.
	Expiry date
LOT	Lot number The number of the kit charge.
IVD	CE-IVD symbol in-vitro diagnostic medical device.
	Manufactured by Contact information of manufacturer.
	For single use only Do not use components for a second time.
	Note / Attention Observe the notes marked in this way to ensure correct function of the kit and to avoid operating errors for obtaining correct results.

The following systematic approach is introduced in the manual:

- The chapters and figures are numbered consecutively.
- A cross reference is indicated with an arrow (e.g. \rightarrow "Notes on the use of this manual" p. 3).
- Working steps are numbered.

2 Safety precautions

NOTE

Read through this chapter carefully prior to guarantee your own safety and a trouble-free operation.

Follow all the safety instructions explained in the manual, as well as all messages and information, which are shown.

All due care and attention should be exercised in handling the materials and reagents contained in the kit. Always wear gloves while handling these reagents and avoid any skin contact! In case of contact, flush eyes or skin with a large amount of water immediately.



FOR SINGLE USE ONLY!

This kit is made for single use only!

ATTENTION!

Don't eat or drink components of the kit!

The kit shall only be handled by educated personnel in a laboratory environment!

If the buffer bottles are damaged or leaking, wear gloves and protective goggles when discarding the bottles in order to avoid any injuries. This kit could be used with potential infectious samples. Therefore, all liquid waste must be considered as potentially infectious and must be handled and discarded according to local safety regulation.

Please observe the federal, state and local safety and environmental regulations. Follow the usual precautions for applications using extracted nucleic acids. All materials and reagents used for DNA or RNA isolation should be free of DNases or RNases.

ATTENTION!

Do not add bleach or acidic components to the waste after sample preparation!

NOTE

Emergency medical information in English and German can be obtained 24 hours a day from:

Poison Information Center, Freiburg / Germany

Phone: +49 (0)761 19 240.

For more information on GHS classification and the Safety Data Sheet (SDS) please dial +49 30 9489 3380.

3 Storage conditions

Store lyophilized **Proteinase K** at 4 °C to 8 °C! Divide dissolved **Proteinase K** into aliquots and storage at -22 °C to -18 °C is recommended. Repeated freezing and thawing will reduce the activity dramatically!

All other components of the deltaPREP Blood DNA Mini Kit (MDX) should be stored dry, at room temperature (15 °C to 30 °C). When stored at room temperature, the kit is stable until the expiration date printed on the label on the kit box.

If there are any precipitates within the provided solutions solve these precipitates by careful warming. Before every use make sure that all components have room temperature.

4 Function testing and technical assistance

The IST Innuscreen GmbH guarantees the correct function of the kit for applications as described in the manual. This kit has been produced and tested in an ISO 13485 certified facility.

We reserve the right to change or modify our products to enhance their performance and design. If you have any questions or problems regarding any aspects of the deltaPREP Blood DNA Mini Kit (MDX), please do not hesitate to contact us. For technical support or further information in Germany please dial +49 30 9489 3380.

5 Product use and warranty

The kit is not designed for the usage of other starting materials or other amounts of starting materials than those, referred to in the manual (→ "Product specifications" p. 8). Since the performance characteristics of our kits have not been validated for any specific application. IST Innuscreen GmbH kits may be used in clinical diagnostic laboratory systems after the laboratory has validated the complete diagnostic system as required by CLIA' 88 regulations in the U.S. or equivalents in other countries.

All products sold by the IST Innuscreen GmbH are subjected to extensive quality control procedures and are warranted to perform as described when used correctly. Any problems should be reported immediately.

NOTE

The kit is an *in-vitro* diagnostic medical product!



6 Kit components

6.1 Included kit components

	Σ 10	Σ 50	Σ 250
REF	31-DP-1000010	31-DP-1000050	31-DP-1000250
Lysis Solution SLS	5 ml	25 ml	120 ml
Binding Solution BL	8 ml	40 ml	200 ml
Proteinase K	for 1 x 0.3 ml working solution	for 2 x 1.5 ml working solution	for 6 x 1.5 ml working solution
Washing Solution C	5 ml	25 ml	120 ml
Washing Solution BS (conc.)	2 ml	8 ml	2 x 18 ml
Elution Buffer	2 x 2 ml	12 ml	3 x 25 ml
Spin Filter	10	50	5 x 50
Receiver Tubes	50	5 x 50	25 x 50
Elution Tubes	10	50	5 x 50
Manual	1	1	1

6.2 Components not included in the kit

- ddH₂O for dissolving Proteinase K
- 1.5 ml tubes
- 96-99.8 % ethanol (molecular biology grade, undenaturated)
- 1 x PBS (137 mM NaCl, 2.7 mM KCl, 10 mM Na₂HPO₄, 1.8 mM KH₂PO₄)
- 2.0 ml tubes; optional
- RNase A (10 mg/ml); optional

7 Product specifications

- 1. Starting material:
- Fresh or frozen whole blood samples
- Stabilizers: EDTA or citrate
- 200 μl sample volume
- 400µl sample volume

NOTE

Avoid freezing and thawing of starting material.

- 2. Time for isolation:
- Approximately 24 minutes including lysis step
- 3. Typical yield:
- Depends on sample (amount of nucleated cells) and amount of starting material
- Up to > 30 μg gDNA
- 4. Typical ration A_{260} : A_{280} :
 - **■** 1.7-2.0

8 Initial steps before starting

- Heat thermal mixer or water bath at 60 °C.
- Add to **Proteinase** K the indicated amount of ddH₂O, mix thoroughly and store as described above.

31-DP-1000010	Add 0.3 ml ddH_2O to lyophilized Proteinase K.	
31-DP-1000050	Add 1.5 ml ddH_2O to lyophilized Proteinase K.	
31-DP-1000250		

Add to Washing Solution BS (conc.) the indicated amount of absolute ethanol, mix thoroughly and store as described above.

31-DP-1000010	Add 18 ml ethanol to 2 ml Washing Solution BS (conc.).
31-DP-1000050	Add 72 ml ethanol to 8 ml Washing Solution BS (conc.).
31-DP-1000250	Add 162 ml ethanol to 18 ml Washing Solution BS (conc.).

- Centrifugation steps should be carried out at room temperature.
- Preheat the Elution Buffer at 60 °C.

9 Protocols for isolation from whole blood samples

9.1 Protocol 1: Isolation from 200 µl blood samples

IMPORTANT

Pre-fill the needed amount of **Elution Buffer** into a 2.0 ml reaction tube and incubate the **Elution Buffer** at 60 °C until the elution step. If the sample volume is less than 200 µl, add the appropriate volume of PBS.

- 1. Pipette **200** µl of whole blood sample into a 1.5 ml reaction tube.
- 2. Add 200 μ I Lysis Solution SLS <u>and</u> 20 μ I Proteinase K, mix vigorously by pulsed vortexing for 10 seconds and incubate the sample at 60 °C for 10 minutes.

NOTE

We recommend using a shaking platform (thermal mixer, water bath or another rocking platform) for a continuous shaking of the sample. Alternatively, vortex the sample 3–4 times during the incubation. No shaking will reduce the lysis efficiency.

NOTE

The kit co-purify DNA and RNA, if DNA and RNA in the sample. If RNA-free genomic DNA is required, add 1-2 μ l of a RNase A stock solution (10 mg/ml) to the sample before addition of **Binding Solution BL**, vortex shortly and incubate for 5 minutes at room temperature.

- 3. Optional: centrifuge the 1.5 ml reaction tube for 10 seconds to remove condensate from the lid of the reaction tube.
- 4. Open the 1.5 ml reaction tube and add 350 μl Binding Solution BL to the lysed sample. Mix carefully by pipetting up and down several times (3 4 times), apply the sample using the pipette to a Spin Filter located in a Receiver Tube and close the cap of the Spin Filter.

IMPORTANT NOTE

Don't vortex the sample at this step!

It is important that the sample and the Binding Solution BL are mixed by pipetting up and down several times. Vortexing will lead to reduced yield of DNA.

5. Centrifuge at 11,000 x g (~12,000 rpm) for 1 minute.
Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.

NOTE

If the solution has not completely passed through the Spin Filter, centrifuge again at higher speed or prolong the centrifugation time.

- 6. Open the Spin Filter and add 400 μ l Washing Solution C, close the cap and centrifuge at 11,000 x g (~12,000 rpm) for 1 minute. Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.
- 7. Open the Spin Filter and add **600 \mul Washing Solution BS**, close the cap and centrifuge at 11,000 x g (~12,000 rpm) for 1 minute. Discard the filtrate and re-use the Receiver Tube. Place the Spin Filter into a new Receiver Tube.
- Open the Spin Filter and add 600 μl Washing Solution BS, close the cap and centrifuge at 11,000 x g (~12,000 rpm) for 1 minute.
 Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.
- 9. Centrifuge at max. speed for 3 minutes to remove all traces of ethanol. Discard the Receiver Tube.
- 10. Place the Spin Filter into an Elution Tube. Carefully open the cap of the Spin Filter and add **200 μl Elution Buffer** (pre-warmed at 60 °C). Incubate at room temperature for 2 minutes.
- 11. Centrifuge at 11,000 x g (~12,000 rpm) for 1 minute. Two elution steps with equal volumes of pre-warmed Elution Buffer (e.g. 100 µl + 100 µl) might increase the yield of extracted gDNA.

NOTE

The DNA can be eluted with a lower or a higher volume of **Elution Buffer** (depends on the expected yield of genomic DNA). Elution with lower volumes of **Elution Buffer** increases the final concentration of DNA. Store the extracted DNA at 4 $^{\circ}$ C to 8 $^{\circ}$ C. For long time storage placing at -22 $^{\circ}$ C to -18 $^{\circ}$ C is recommended.

9.2 Protocol 2: Isolation from 400 µl blood samples

IMPORTANT NOTE

Pre-fill the needed amount of Elution Buffer into a 2.0 ml reaction tube and incubate the Elution Buffer at 60 °C until the elution step.

- 1. Pipette 400 µl of whole blood sample into a 2.0 ml reaction tube.
- 2. Add **400 μl Lysis Solution SLS** <u>and</u> **30 μl Proteinase K**, mix vigorously by pulsed vortexing for 10 seconds and incubate the sample at 60 °C for 10 minutes.

NOTE

We recommend using a shaking platform (thermal mixer, water bath or another rocking platform) for a continuous shaking of the sample. Alternatively, vortex the sample 3–4 times during the incubation. No shaking will reduce the lysis efficiency.

NOTE

The kit co-purifies DNA and RNA, if DNA and RNA in the sample. If RNA-free genomic DNA is required, add 1-2 μ l of a RNase A stock solution (10 mg/ml) to the sample before addition of **Binding Solution BL**, vortex shortly and incubate for 5 minutes at room temperature.

- 3. Optional: centrifuge the 1.5 ml reaction tube for 10 seconds to remove condensate from the lid of the reaction tube.
- 4. Open the 1.5 ml reaction tube and add 700 μl Binding Solution BL to the lysed sample. Mix carefully by pipetting up and down several times (3 4 times), apply 750 μl of the sample using the pipette to a Spin Filter located in a Receiver Tube and close the cap of the Spin Filter.

IMPORTANT NOTE

Don't vortex the sample at this step!

It is important that the sample and the Binding Solution BL are mixed by pipetting up and down several times. Vortexing will lead to reduced yield of DNA.

5. Centrifuge at 11,000 x g (~12,000 rpm) for 1 minute.

Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.

NOTE

If the solution has not completely passed through the Spin Filter, centrifuge again at higher speed or prolong the centrifugation time.

- 6. Apply the residual sample to the Spin Filter. Close the cap and centrifuge at $11,000 \times g$ (~12,000 rpm) for 1 minute.
 - Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.
- 7. Open the Spin Filter and add 400 μ l Washing Solution C, close the cap and centrifuge at 11,000 x g (~12,000 rpm) for 1 minute. Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.
- Open the Spin Filter and add 600 μl Washing Solution BS, close the cap and centrifuge at 11,000 x g (~12,000 rpm) for 1 minute.
 Discard the filtrate and re-use the Receiver Tube. Place the Spin Filter into a new Receiver Tube.
- Open the Spin Filter and add 600 μl Washing Solution BS, close the cap and centrifuge at 11,000 x g (~12,000 rpm) for 1 minute.
 Discard the Receiver Tube with the filtrate. Place the Spin Filter into a new Receiver Tube.
- 10. Centrifuge at max. speed for 3 minutes to remove all traces of ethanol. Discard the Receiver Tube.
- 11. Place the Spin Filter into an Elution Tube. Carefully open the cap of the Spin Filter and add **200 μl Elution Buffer** (pre-warmed at 60 °C). Incubate at room temperature for 2 minutes.
- 12. Centrifuge at $11,000 \times g$ (~12,000 rpm) for 1 minute. Two elution steps with equal volumes of pre-warmed Elution Buffer (e.g. 100 μ l + 100 μ l) might increase the yield of extracted gDNA.

NOTE

The DNA can be eluted with a lower or a higher volume of Elution Buffer (depends on the expected yield of genomic DNA). Elution with lower volumes of Elution Buffer increases the final concentration of DNA. Store the extracted DNA at 4 $^{\circ}$ C to 8 $^{\circ}$ C. For long time storage placing at -22 $^{\circ}$ C to -18 $^{\circ}$ C is recommended.

10 Troubleshooting

Problem / probable cause	Comments and suggestions			
Clogged Spin Filter				
Insufficient disruption or homoge-	Increase lysis time.			
nization	Increase centrifugation speed.			
	Reduce amount of starting material.			
Little or no DNA eluted				
Insufficient lysis	Increase lysis time.			
	Reduce amount of starting material. Over-			
	loading of Spin Filter reduces yield!			
Incomplete elution	Prolong the incubation time with Elution			
	Buffer to 5 minutes or repeat elution step			
	once again.			
	Take a higher volume of Elution Buffer.			
Insufficient mixing with Binding	Mix sample with Binding Solution BL by pi-			
Solution BL	petting up and down several times prior to			
	transfer of the sample onto the Spin Filter.			
Low concentration of extracted				
DNA				
Too much Elution Buffer	Elute the DNA with lower volume of Elution			
	Buffer.			
Degraded or sheared DNA				
Incorrect storage of starting mate-	Ensure that the starting material is frozen			
rial	immediately in liquid N_2 or in minimum at			
	20° C and is stored continuously at -80° C!			
	Avoid thawing of the material.			
Old material insufficient	Old material often contains degraded DNA.			
RNA contaminations of extracted DNA				
RNA contaminations of extracted	Perform RNase digestion			
DNA				

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