## Enrichment of Biomolecules

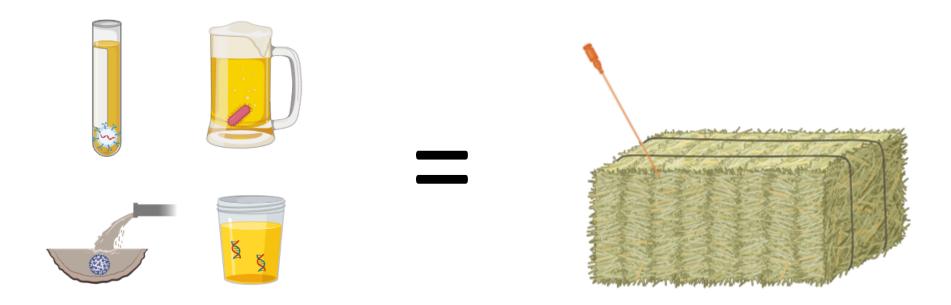
Dr. Sandra Tückmantel

# ST Innuscreen GmbH and Berlin



### Why target enrichment ?



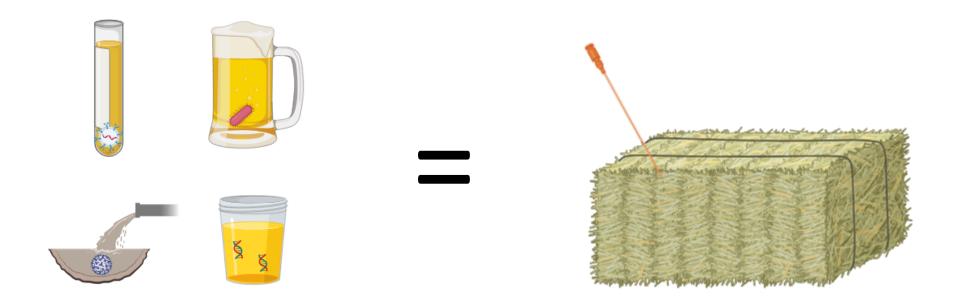


large sample volume + few target molecules = needle in the haystack

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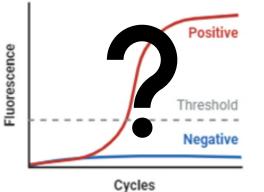
## Why target enrichment?





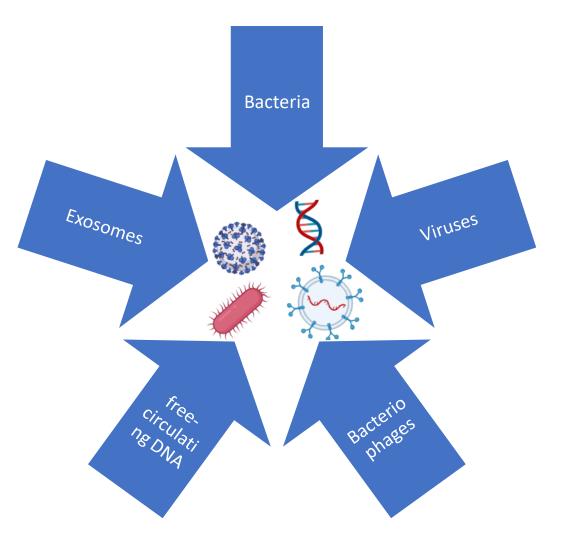
large sample volume + few target molecules = needle in the haystack

direct DNA/RNA extraction + PCR often not possible!



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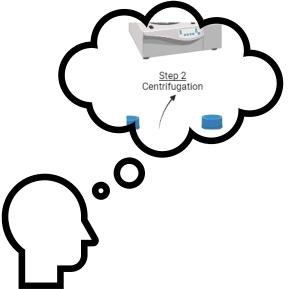
### What are the targets of interest?



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Centrifugation?

- good for bacteria, but ultra-centrifugation necessary for viruses, cell-free DNA, exosomes...
- difficult for high sample volumes

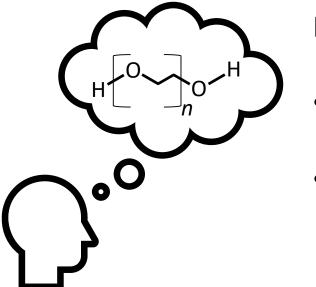




Filtration?

- risk of filter clogging
- sample loss if pore size too large





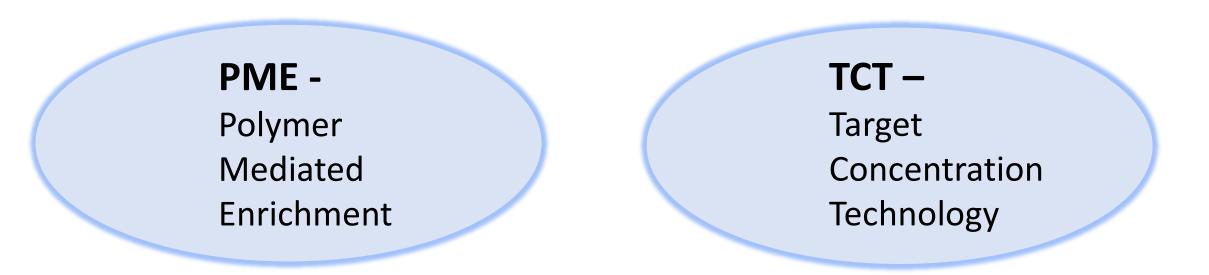
**PEG Precipitation?** 

- time-consuming
- labor-intensive

# New enrichment technologies by IST Innuscreen IST GmbH

### **Our Solution:**

Two new technologies for enrichment of biomolecules for sample pre-treatment of aqueous liquids (water, beverages, body fluids).





**General principle of PME** 



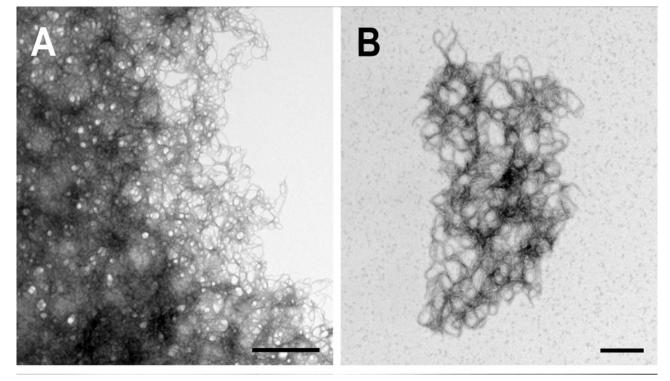
Sample + Reagent 1 and 2 – formation of biomolecule/polymer-complex Centrifugation of biomolecule/polymer-complex

Dissolving of biomolecule/polymer-complex (e.g. in lysis buffer for DNA isolation)

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### Polymer Mediated Enrichment (PME) – Enrichment of virus particle



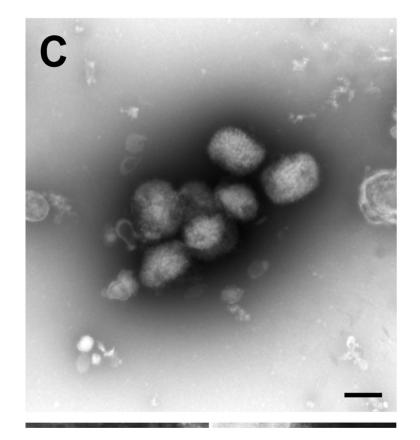
Electron micrograph of polymer-structure

A-B: Electron micrograph of polymer complex; Bars: A = 500 nm, B = 200 nm

> (data kindly provided by P. Patel; RKI Berlin)



### Polymer Mediated Enrichment (PME) – Enrichment of virus particle



Electron micrograph of Camelpox Virus

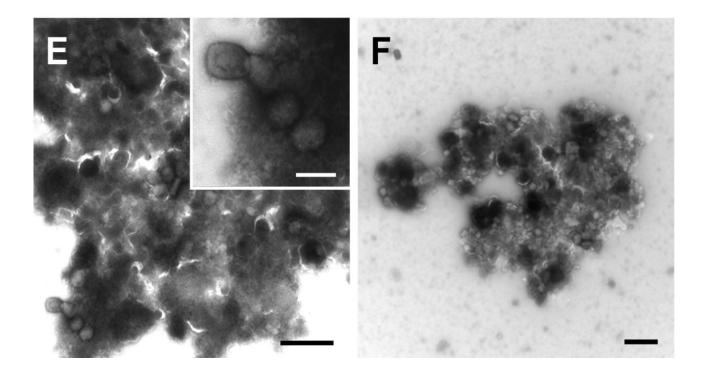
C: Electron micrograph of Camelpox Virus

Bars: C = 200 nm, D = 200 nm

> (data kindly provided by P. Patel; RKI Berlin)



### Polymer Mediated Enrichment (PME) – Enrichment of virus particle



Electron micrograph of polymer- Camelpox virus

E-F: Electron micrograph of polymer-virus complex after enrichment Bars: E = 1  $\mu$ m F = 1  $\mu$ m

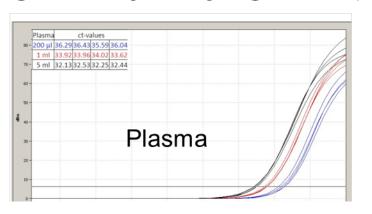
(data kindly provided by P. Patel; RKI Berlin)

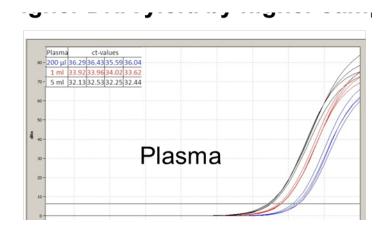
**Innuscreen** GmbH

Sometimes more is more....

Extraction of cell-free DNA followed by qPCR detection (human-specific sequence) with

- 200 µl (standard spin filter extraction)
- 1 ml (enriched using PME Technology)
- 5 ml (enriched using PME Technology)
- 10 ml (enriched using PME Technology)





### Higher DNA yield by higher sample $\boldsymbol{\nu}$

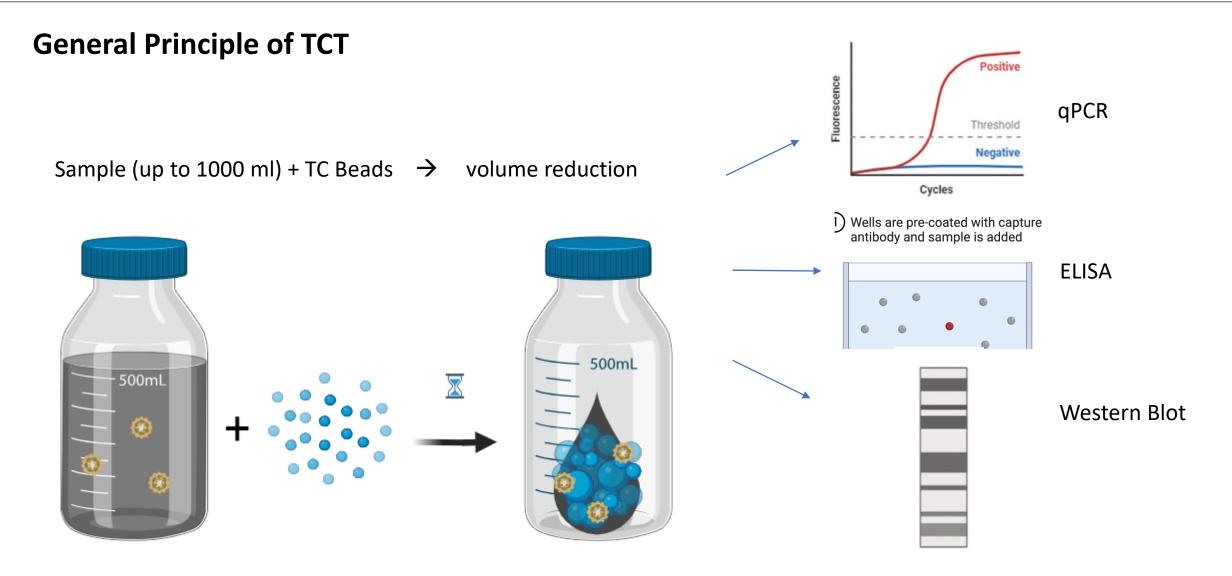


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### Summary PME Technology

- suitable for water, cell culture supernatant, serum, plasma, liquor, urine...
- for sample volumes of up to 50 ml
- enrichment of viruses, virus fragments, free-circulating DNA, bacteriophages, bacteria, exosomes
- enriched viruses suitable for re-cultivation, cell assays etc. (no harmful effects by polymer)



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**Example: concentration of bacteriophages in water samples** 

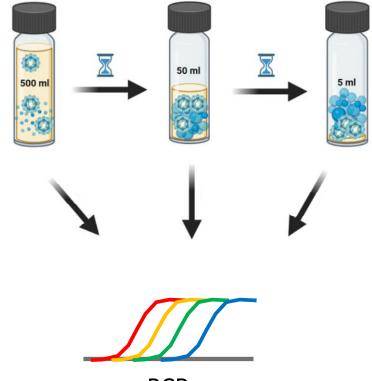
Sample: 500 ml drinking water

Spike: bacteriophage MS2

Procedure: addition of TCT beads to the sample and target enrichment in 2 steps:

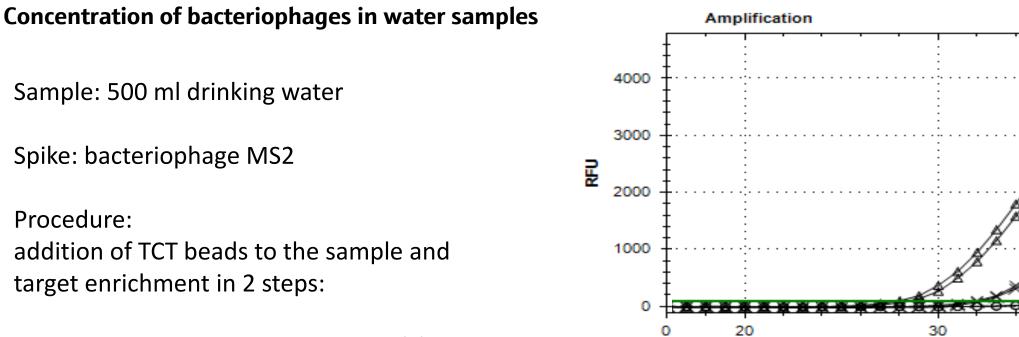
Step 1 – volume reduction to 50 ml Step 2 – volume reduction to 5 ml

Extraction of MS2-RNA from 200  $\mu$ l of each sample and subsequent RT-PCR for detection



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**qPCR** parts of scheme created with app.biorender.com



Step 1 -volume reduction to 50 ml(x)Step 2 – volume reduction to 5 ml ( $\triangle$ )

Procedure:

Extraction of MS2-RNA from 200 µl of each sample and subsequent RT-PCR for detection

sample	symbol	CT value	enhancing factor
initial sample 1 (200 μl of 500 ml)	0	35,93	0
initial sample 2 (200 μl of 500 ml)	0	36,73	0
sample 1 from 1. enhancing step (200 μl of 50 ml)	Х	31,72	10
sample 2 from 1. enhancing step (200 μl of 50 ml)	Х	32,07	10
sample 1 from 2. enhancing step (200 μl of 5 ml)	Δ	28,23	100
sample 2 from 2. enhancing step (200 μl of 5 ml)	Δ	27,68	100

Cycles

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Detection of low concentrations of free circulating plasmid DNA

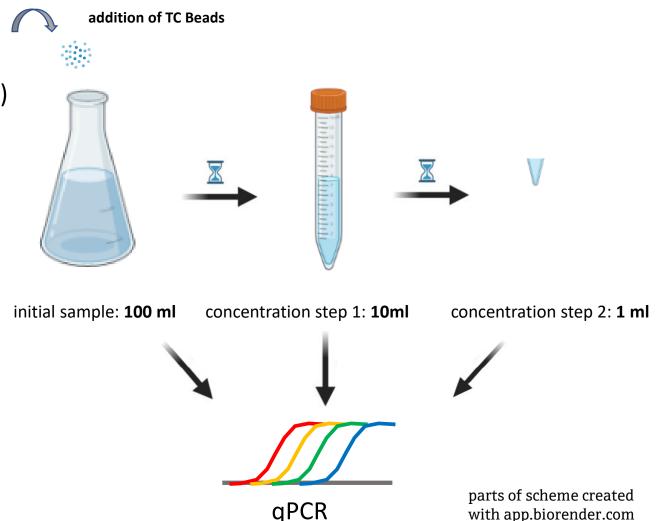
Sample: 100 ml drinking water

Spike: 100 fg plasmid DNA (1 fg/ml = 580 molecules/ml)

Procedure: addition of TCT beads to the sample and target enrichment in 2 steps:

Step 1 – volume reduction to 10 ml Step 2 – volume reduction to 1 ml

Extraction of plasmid DNA and subsequent RT-PCR for detection



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### **Detection of low concentrations of free circulating plasmid DNA**

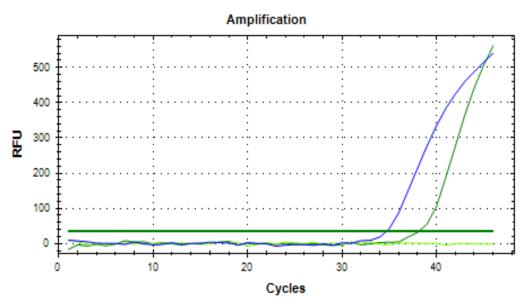
Sample: 100 ml drinking water

Spike: 100 fg plasmid DNA (1 fg/ml = 580 molecules/ml)

Procedure: addition of TCT beads to the sample and target enrichment in 2 steps:

Step 1 – volume reduction to 10 ml Step 2 – volume reduction to 1 ml

Extraction of plasmid DNA and subsequent RT-PCR for detection



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Sample	color	Ct value	volume	concentration factor
1	light green	no detection	100 ml	
2	green	38,1	10 ml	10
3	blue	34,8	1 ml	100

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GmbH

### **Enrichment of bacteria from a bacterial culture**

Sample: E.coli

Samples volume: 50 ml growth medium

Procedure: addition of TCT Beads to 50 ml of bacterial culture and concentration from 50 ml to 10 ml

Plating out on agar plates and incubation

### **Enrichment of bacteria from a bacterial culture**

Sample: E.coli

Samples volume: 50 ml growth medium

Procedure: addition of TCT Beads to 50 ml of bacterial culture and concentration from 50 ml to 10 ml

Plating out on agar plates and incubation

with enrichment (factor 5)

no enrichment





### Summary TCT

- for sample volumes of up to 1000 ml
- for enrichment of viruses, virus fragments, free-circulating NA, bacteriophages, bacteria, proteins, etc.
- enriched sample can be directly used for downstream applications

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- compatible with any kind of DNA/RNA extraction method
- enrichment of living bacteria possible

# Thank you for your attention!